Staining Intensity of P16\textsuperscript{INK4a} and Ki-67 Determine the Grade of Cervical Lesion: An Experience from Single Tertiary Care Centre

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Abstract

**Background:** Various non-neoplastic and neoplastic lesions occur in cervix. HPV infection plays a major role in genesis of cervical lesions. Although most HPV infections are cleared out, infections with high risk HPVs may persist resulting in cervical dysplastic lesions. Using p16, a surrogate marker for HPV infection and Ki-67, a proliferation marker, along with histopathology, help improve the diagnostic accuracy of these lesions. The aim of the study was to evaluate the expression of p16 and Ki-67 in cervical lesions and association of their staining intensity with the histologic grading. Also to determine the predictive value of these markers in association with the cervical lesions. **Methodology:** A cross sectional study was carried out in 122 cases which were diagnosed histopathologically and then evaluated for the immunohistochemical expression of p16 and Ki-67 and their staining intensities. **Results:** The dysplastic lesions comprised of 83.7% cases [29.5% LSIL, 5.7% HSIL, 43.5% SCC and 4.9% adenocarcinoma] and 16.3% had non dysplastic lesions. p16 and Ki-67 expression were seen in 64.7% and 68% cases respectively. The intensity of p16 and Ki-67 expression was scored according to Galgano et al (2010). An increasing intensity of p16 and Ki-67 expression with higher grades of the cervical lesions was noted and this association was found to be statistically significant. ($\chi^2 = 43.46$ and p value < 0.0001). **Conclusion:** Though histopathology is the gold standard, the role of p16 and Ki-67 have emerged as useful adjuncts in detecting the true nature of the cervical lesions. They aid in the proper diagnosis, classification and distinction from non-dysplastic lesions, helping the clinicians in taking prompt action for management of the cases.

Keywords: Squamous Intraepithelial Lesion- cervical carcinoma- immunohistochemistry- p16 and Ki-67

Introduction

The lesions encountered in cervix vary from mere non neoplastic inflammatory lesions to dysplastic lesions, Squamous Intraepithelial Lesion and carcinoma of uterine cervix and they occur mainly due to sexual and behavioural factors, socio-economic factors (education and income), smoking, diet, oral contraceptives, hormones, other infections like herpes simplex virus (HSV), human immunodeficiency virus (HIV) [1]. According to Walboomers (1999) infection with Human Papilloma Virus (HPV), mainly subtypes 16 and 18 are implicated in cervical neoplasms [2,3]. The imbalance between persistence of HPV infections and clearing it off results in most HPV associated cervical lesions. Most HPV infections are self limiting or regress if treated early. However, infection with high-risk HPVs may progress into much severe forms like dysplasia, Squamous Intraepithelial Lesion and later into cervical cancer if the HPV infection persists. But carcinoma cervix can be 100% preventable as its rate of progression is very slow with a long preinvasive stage. Also, with the presence of diagnostic procedures like cervical cytology and histopathology and use of several biomarkers, prompt early diagnosis makes it readily treatable giving the patients a long survival period [4].

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Also, cervical cancer can be prevented by use of vaccines. Two vaccines licensed globally are available in India against cervical cancer. Cervarix, a bivalent vaccine against serotypes 16 & 18 and Gardasil, a quadrivalent vaccine against serotypes 16, 18, 6 & 11. The recommended age to initiate vaccination is 9–12 years. Catch-up vaccination can be done up to the age of 26 years. A total of three doses at 0, 2 and 6 months are recommended for Gardasil or 0, 1 and 6 months for Cervarix. The vaccines provide protection against cervical cancer and genital warts as well [5].

Cancer of uterine cervix is the fourth most common cancer recorded in women worldwide. It is also the fourth leading cause of cancer deaths among women worldwide with an estimated 570,000 cases and 311,000 deaths in 2018. In lower Human Development Index setting, cervical cancer ranks as second in both incidence and mortality [6]. Cervical cancer accounts for 17% of all cancer deaths amongst women aged 30-69 years of age [7,8]. Poor living standards, a higher prevalence of HPV (10% in women above the age of 30 years) and due to lack of screening are the causes of cervical cancer [8].

In Indian scenario, carcinoma of cervix is the second most common cancer in women [6]. India has a burden of cervical carcinoma accounting nearly 1/3rd of global carcinoma of cervix [9]. Among the women of lower socio-economic status, the burden of cervical cancer is much higher. It has been reported that every 7 minutes an Indian woman dies of cervical cancer [10].

p16INK4a is a tumor-suppressor protein. It works by blocking cyclin dependent kinases 4/6-mediated pRb phosphorylation and inhibits E2F-dependent transcription and cell-cycle progression [11]. The E7 protein of high-risk HPVs inactivates pRB, hence there is resulting overexpression of p16INK4a which may be used as a good marker for infection with high risk HPV types. Immunostaining of p16INK4a allows precise identification of even small CIN or cervical cancer lesions in biopsy sections. Thus it can help reduce inter-observer variation in the histopathological interpretation of cervical biopsy specimens [12,13].

Ki67 is used as an indicator of cellular proliferation frequently. Ki67 is expressed more significantly in malignant than in normal tissues. Ki67 increases with decreasing tissue differentiation, and it correlates with the clinical stage of tumors and presence of occult metastasis. [14,15].

The p16/Ki-67 co-expression implies deregulation of the cell cycle induced by HR-HPV and detection of p16/Ki-67 co-expression can serve as a marker to predict the cell transformation by HR-HPV and the presence of high-grade CIN lesions [16].

As the cell cycle controls are overruled by HPV infections, the detection of cell proteins p16 and ki67 which are differentially expressed in HPV infected cells are currently being considered for cervical cancer screening and as prognostic markers [13].

The study was carried out to evaluate the expression of p16 and Ki-67 in cervical lesions and to see the association of the staining intensity of the markers with the histologic grading. Also to determine the predictive value of these markers in association with the cervical lesions.

**Materials and Methods**

A hospital based cross sectional study was conducted for a period of one year from July 2021 to June 2022 in the Department of Pathology in collaboration with Department of Obstetrics and Gynaecology, GMCH. The approval was given by Institutional ethics committee having vide letter no. 190/2007/Pt-II/July-2021/TH-10.

**Inclusion Criteria**

Cervical biopsies and surgically resected specimens of cervix sent to the Department of Pathology, Gauhati Medical College and Hospital.

**Exclusion Criteria**

1. Inadequate biopsy, ulcerated and necrotic tissue.
2. Endocervical polyps.
5. Patients who did not give consent.

A total of 122 cases were collected following the inclusion and exclusion criteria from Gynaecology OPD who presented with complaints of bleeding PV, white discharge, irregular menses, etc and underwent biopsy and hysterectomy. All the demographic as well as the clinical data were entered in MS Excel sheet.

Nature of specimen received were: Punch Biopsy: 102, Amputated cervix: 1, Hysterectomy: 18, Modified Radical hysterectomy: 1. Specimens were collected in 10% Neutral Buffered Formalin and the specimens were grossed, processed and then stained as per standard protocols [17]. Diagnosis was made based on the morphologic changes noted in the lining epithelium of the cervix along with the stromal reaction. For reporting, we followed the standard pathology textbook (Rosai and Ackerman’s surgical pathology) [18]. For tumors, we followed WHO classification of Tumours of Female Reproductive Organs- Tumours of the uterine cervix (2022) [19]. The cases were then evaluated for p16 and Ki67 immunostain on representative blocks of paraffin embedded tissue. Immunohistochemistry procedure was carried out according to standard protocols as follows: [17].

All the paraffin-embedded tissue blocks were cut at 4 μM thickness. Two sections were made ready from each block of which one was placed on albuminized slides for routine H&E stain and the other set was mounted on poly L-lysine coated slides for immunostain. The poly L-lysine coated slides were placed in the oven for 10 minutes after which deparaffinization was done by passing the sections through xylene. Subsequently, sections were rehydrated in graded alcohol of decreasing concentration i.e. 100%, 70% and 50% for five minutes each. The sections were then rinsed in running water for five minutes. Antigen retrieval was performed using the microwave method (also known as Heat Induced Epitope Retrieval, HIER) is used in our IHC set up, where, the slides were immersed in a bowl containing TRIS-EDTA buffer (6.05g of TRIS and
Statistical Analysis

The data collected was analyzed using SPSS version 20. The association between histopathological diagnosis and each biomarker was statistically calculated by using the Chi-square ($\chi^2$) test of significance by adopting the statistical software SPSS. P values less than 0.5 was considered statistically significant. Sensitivity, specificity, and predictive values were calculated using $2 \times 2$ tables and standard formula.

Results

Histological diagnosis of the 122 cases were as follows: 43 cases had squamous intraepithelial lesions of which 29.5% (36/43 cases) had LSIL and 5.7% (7/43 cases) had HSIL, 59 cases had cervical cancer of which 43.5% (53/59 cases) were squamous cell carcinoma and 4.9% (6/59 cases) had adenocarcinoma. 20 cases were non-dysplastic of which 9.8% (12/20) cases had chronic cervicitis and 6.6% (8/20) cases had metaplastic change.

It was found in our study that 79/122 cases (64.7%) showed p16 immunoexpression whereas 43/122 cases (35.3%) were negative for p16. It was also found that p16 immunoexpression was highest in cervical squamous cell carcinoma [55/122 cases (45.1%)], followed by 22/122 cases of SIL and 2/122 cases of non-dysplastic lesions, which indicated that the p16 expression increased with the increasing grades of the cervical lesion. A significant association was observed between routine histologic grading and p16 immunoexpression, Chi-square value is 50.70 and p value is <0.001.

The intensity of p16 expression was scored according to Galgano et al [20]. Among the 20 non-dysplastic lesions, 2 metaplastic lesions showed mild (grade 1) p16 positivity. Out of 15/36 LSIL cases that were p16 positive, 8/15 cases showed grade 1, 6/15 cases showed grade 2 and 1/15 cases showed grade 3 positivity. All the 7 cases of HSIL, 51/53 cases of cervical squamous cell carcinoma and 4/6 cervical adenocarcinoma cases showed p16 positivity. Of the 7 HSIL cases, 1/7 cases showed grade 1, 2/7 cases showed grade 2 and 4/7 cases showed grade 3 positivity for p16. Of the cervical SCC cases, 51/53 showed p16 positivity of which 2/51 cases showed grade 1, 4/51 cases showed grade 2 and 45/51 cases showed grade 3 positivity.

Of the 4/6 cervical adenocarcinoma cases that were positive for p16, 1/4 cases showed grade 2 positivity while 3/4 cases showed grade 3 positivity. An increasing trend of p16 expression intensity from focal positivity in low grade lesions to diffuse intensity in higher grade lesions was found which was statistically significant. [chi square value = 76.87 and p value <0.0001] (Table 1). Similarly, 83/122 cases (68%) were positive for Ki-67 and 39/122 cases (32%) were negative for Ki-67
immunoexpression. We also observed that Ki-67 immunostaining was highest in cervical carcinoma cases, 56/122 cases (46%) followed by 25/122 cases (20.4%) of SIL and 2/122 cases (1.6%) of non-dysplastic lesions, which indicated that the ki67 expression increased with the increasing grades of the cervical lesion. A significant association was observed between routine histologic grading and Ki-67 immunoexpression, Chi square value is 52.51 and p value is <0.001.

The intensity of ki67 expression was scored according to Galgano et al. [20] of the 18/36 LSIL cases positive for ki67, 12/18 cases had score 1 and 6/18 cases had score 2. Of the 7/7 HSIL cases positive for KI67, 1/7 case had score 1, 3/7 cases had score 2 and 3/7 cases had score 3. Of the 51/53 SCC cases positive for KI67, 1/5 had score 2, 20/51 cases had score 2 and 26/51 cases had score 3. Of the 5/6 adenocarcinoma cases positive for ki67, 1/5 cases had a score 1, 1/5 cases had score 2 and 3/5 cases had score 3 (Table 2).

Table 1. Association of p16 Expression with Grade of Cervical Lesion (n=122)

<table>
<thead>
<tr>
<th>Cervicitis</th>
<th>Metaplasia</th>
<th>LSIL</th>
<th>HSIL</th>
<th>SCC</th>
<th>Adeno Ca</th>
<th>Total cases (%)</th>
<th>d.f</th>
<th>Chi square value</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative (Grade 0)</td>
<td>12</td>
<td>6</td>
<td>21</td>
<td>0</td>
<td>2</td>
<td>2</td>
<td>43 (35.2)</td>
<td>15</td>
<td>76.87</td>
</tr>
<tr>
<td>Mild Positivity (Grade 1)</td>
<td>0</td>
<td>2</td>
<td>8</td>
<td>1</td>
<td>2</td>
<td>0</td>
<td>13 (10.7)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Moderate Positivity (Grade 2)</td>
<td>0</td>
<td>0</td>
<td>6</td>
<td>2</td>
<td>4</td>
<td>1</td>
<td>13 (10.7)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intense Positivity (Grade 3)</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>4</td>
<td>45</td>
<td>3</td>
<td>53 (43.4)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>12</td>
<td>8</td>
<td>36</td>
<td>7</td>
<td>53</td>
<td>6</td>
<td>122 (100)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 2. Association of Intensity of ki67 Stain with Cervical Lesion Grading (n=122)

<table>
<thead>
<tr>
<th>Score 0</th>
<th>Score 1</th>
<th>Score 2</th>
<th>Score 3</th>
<th>Total cases (%)</th>
<th>d.f</th>
<th>Chi square</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Score 0</td>
<td>12</td>
<td>6</td>
<td>18</td>
<td>0</td>
<td>2</td>
<td>1</td>
<td>39 (32)</td>
</tr>
<tr>
<td>Score 1</td>
<td>0</td>
<td>1</td>
<td>12</td>
<td>1</td>
<td>5</td>
<td>1</td>
<td>20 (16.4)</td>
</tr>
<tr>
<td>Score 2</td>
<td>0</td>
<td>1</td>
<td>6</td>
<td>3</td>
<td>20</td>
<td>1</td>
<td>31 (25.4)</td>
</tr>
<tr>
<td>Score 3</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>3</td>
<td>26</td>
<td>3</td>
<td>32 (26.2)</td>
</tr>
</tbody>
</table>

[p16 and 79.41%, 90%, 97.59% and 46.15% respectively for Ki-67. The sensitivity and specificity of both stains together is 98.21% and 100% respectively (Table 3), (Figure 1 and 2).

Discussion

A total number of 122 histopathologically diagnosed cases of cervical lesions underwent immunohistochemical examination for p16 and Ki67 in the Department of Pathology. The results and observations are compared with observations of the literatures by other workers in similar studies from different regions of the world with a detailed discussion.

The present study aimed to study p16 and Ki-67 expression and their staining intensity in cervical lesions. It was found in several studies that the normal cervical epithelium did not express p16, but intensity of p16 and Ki-67 expression increased with degree of histologic atypia in the cervical lesions. Diffuse and strong p16 and Ki-67 expression is observed in high grade dysplastic lesions. Aslani et al (2013) recommended the use of these two markers as complementary tests for differentiating between dysplastic and non-dysplastic lesions [21]. In our study as well, the markers were positive in dysplastic lesions but negative in most non dysplastic lesions.

Our study showed that along with the high grade
Table 3. Diagnostic Values of p16 and Ki-67 Expression in Cervical Biopsy

<table>
<thead>
<tr>
<th>Variables</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>Positive predictive value (%)</th>
<th>Negative predictive value (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>p16 and routine histology</td>
<td>75.49 (65.98 to 83.47)</td>
<td>90.00 (68.30 to 98.77)</td>
<td>97.47 (91.14 to 99.31)</td>
<td>41.86 (33.20 to 51.05)</td>
</tr>
<tr>
<td>Ki-67 and routine histology</td>
<td>79.41% (70.27 to 86.78%)</td>
<td>90.00 (68.30 to 98.77%)</td>
<td>97.59 (91.55 to 99.34%)</td>
<td>46.15 (36.30 to 56.32%)</td>
</tr>
<tr>
<td>p16 and Ki-67</td>
<td>98.21% (90.45 to 99.95%)</td>
<td>100% (29.24 to 100%)</td>
<td>75% (29.24 to 100%)</td>
<td>75% (30.07 to 95.44%)</td>
</tr>
</tbody>
</table>

Figure 2. Diffuse Staining Pattern of Ki-67 (score 3) in Cervical Cancer (40X)

cervical lesions, p16 and Ki-67 were positive in CIN [I, II, III] as well. Majority of CIN-I lesions and a few of CIN-II lesions have the capacity to regress spontaneously. Therefore, predicting the development of CIN is also an important issue for cervical cancer prevention and treatment [22].

p16/Ki-67 coexpression showed a strong association with CIN II+ lesions which is due to persistence of HPV infections, especially with HPV16/18. Hence, p16/Ki-67 is considered as a suitable biomarker for cervical cancer screening [23,24]. In our study, we found that diagnosis using p16 has high specificity (90.0%), but the sensitivity is poor (75.49%). But when combined with Ki-67, sensitivity (98.21%) and specificity (100%) were both at a high level.

The combined use of p16 and Ki-67 immunostain can be used as an auxiliary tool in diagnosis of carcinomas of cervix as seen in the present study and in other studies like Quin Shi et al., 2019 [25] Ding et al. in their study, provided an insight in using these markers to identify CIN patients, who are at a higher risk of malignant progression, facilitating more prompt and cost-effective, efficient interventions [26].

Studies by Klaes et al [27], Agoff et al [28], Kanthiya et al [29], Izadi et al [30] and Sarma et al [24] reported an increase in p16 and Ki-67 expression along with increased staining intensity with higher grades of cervical lesions. In the present study as well, similar findings were noted.

The present study aimed to evaluate the expression of the markers p16 and Ki-67 in cervical lesions. An ascending pattern of p16 and Ki-67 immunoreexpression was seen with increasing grades of cervical lesion and it was statistically significant. Also, their staining intensity showed an increasing score with higher grades of the cervical lesions and it was statistically significant. Thus it can be said that p16 and Ki-67 over expression can be associated with dysplastic or neoplastic lesions.

In conclusion, though histopathology is the gold standard, the role of p16 and Ki-67 have emerged as useful adjuncts in predicting the true nature of the cervical lesions. The staining pattern of p16 and Ki-67 in different histologic grades of cervical lesions have justified its usefulness in confirmation of a histologic diagnosis and its biological behaviour. Positive p16 immunostain of the low grade dysplastic lesions may predict their aggressive behaviour and their propensity to turn to a higher grade or malignant lesion. Ki-67 being a proliferation marker, helps in detecting the proliferation potential of the lesions, but it can show positivity in reparative and inflammatory conditions as well. Hence it is imperative to use Ki-67 along with p16, as complementary markers, to differentiate between the dysplastic and non-dysplastic cervical lesions. Thus, the present study establishes the importance of using p16 and Ki-67 immunostains in routine histopathology to detect the true nature of the cervical lesions.

Strength of the Study
1. Data generated from the study has evaluated the utility of the biomarkers p16 and Ki-67 in predicting the nature of the cervical lesions.
2. There is a future scope of molecular or genetic study from the samples that were included in the study. HPV status can be determined in the cases that stained positive for p16 in our study. It may detect the persistence of HPV infection and help in successful management of the cases.

Limitations of the Study
1. A smaller sample size and shorter duration of the study.
2. It was a cross sectional study, hence the cases could not be followed up. The expression of only two markers was studied. Application of more markers would have been beneficial to predict the other parameters of cervical cancer [CD31, CD105, Factor VIII, E-cadherin, P53, BCL2 etc]

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