DOI:10.31557/APJCC.2023.8.4.691

RESEARCH ARTICLE

Bloodstream Infections in Pediatric Cancer Patients with Febrile Neutropenia at a Tertiary Cancer Center in Northeast India

Amrita Talukdar¹, Rashmisnata Barman¹, Munlima Hazarika², Gaurav Das³

¹Department of Microbiology, Dr. Bhubaneswar Borooah Cancer Institute, India. ²Division of Paediatric Medical Oncology, Department of Medical Oncology, Dr. Bhubaneswar Borooah Cancer Institute, India. ³Department of Surgical Oncology, Dr. Bhubaneswar Borooah Cancer Institute, India.

Abstract

Objective: This study aimed to investigate the pattern of microbial flora, their susceptibility patterns, and clinical variables associated with bloodstream infections in febrile neutropenic patients with solid tumors and hematological malignancies in the pediatric age group in Northeast India. **Materials and Methods:** This retrospective, observational study was conducted at a single tertiary care cancer center in Northeast India between January 1, 2020, and December 31, 2021. The study population included all patients under 18 years of age who developed febrile neutropenia during cancer treatment. **Results:** A total of 378 blood culture samples were analyzed. Febrile neutropenia was found in 252 patients (66.7%). Of these, 45 blood cultures were positive (17.8%). Gram-negative and gram-positive organisms accounted for 62% and 38% of all positive cultures, respectively. *Escherichia coli* (39%) was the most common gram-negative isolate, followed by *Klebsiella pneumoniae* (32%), *Pseudomonas aeruginosa* (18%), and *Acinetobacter baumannii* (7%). Coagulase-negative Staphylococci (CoNS) was the most common gram-negative to beta-lactam/beta-lactamase inhibitor (BL/BLI) antibiotics, such as cefaperazone/sulbactam, was observed in 60% of *Pseudomonas isolates*. Sensitivity to colistin was noted in 89% of *Klebsiella* and 82% of *E. coli* isolates. The incidence of methicillin-resistant *Staphylococcus aureus* (MRSA) was 50%. **Conclusion:** Understanding the microbiological profile and resistance patterns among pediatric cancer patients with febrile neutropenia is crucial for establishing effective antimicrobial policies.

Keywords: Febrile neutropenia- paediatric cancer- antimicrobial resistance- haematological malignancies

Asian Pac J Cancer Care, 8 (4), 691-695

Submission Date: 06/29/2023 Acceptance Date: 09/05/2023

Introduction

Febrile neutropenia is a significant cause of morbidity and mortality among cancer patients in the paediatric age group. Bacteremia is the cause of the febrile neutropenia in about one-fourth of the patients. A high mortality rate has been reported in published literature, especially in those who develop septic shock and pneumonia [1, 2]. Despite the advances in treatment protocols including effective empirical broad-spectrum antibiotics, antifungals, and granulocyte colony-stimulating factors, febrile neutropenia remains a therapeutic challenge. It prolongs hospital stay, increases health-care costs, and compromises chemotherapy delivery with delays and dose reductions.

The aim of the present study was to study the pattern of

microbial flora, their susceptibility patterns, and clinical variables among bloodstream infections in febrile neutropenic patients with solid tumors and hematological malignancies in the paediatric age group in North-East India.

Materials and Methods

It was a retrospective and observational study done in a single tertiary care cancer centre in North-East India. The study period was from 1st January 2020 to 31st December 2021. The study population included all the patients below the age of 18 years who developed

Corresponding Author: Dr. Gaurav Das Department of Surgical Oncology, Dr. Bhubaneswar Borooah Cancer Institute, India. Email: das.drgaurav@gmail.com apjcc.waocp.com

febrile neutropenia during treatment for a diagnosed cancer. Data was collected from the patient case records, hospital electronic medical record system and the registers maintained in the Department of Microbiology. The patient's identifiable information details were de-identified upon entry into the case record forms. The study was approved by the institutional ethics committee (IEC) with waiver of consent.

Blood cultures and bacteremia

Blood cultures were done when the patient developed fever and blood investigations were checked at the same time. A blood culture was deemed positive when one or more samples showed the presence of an organism. The exception was in the case of coagulase negative Staphylococci (CoNS), and in such cases, two separate positive blood cultures were necessitated for representing a true result. We studied samples of peripheral blood as well as blood drawn through central venous catheters (CVCs), peripherally inserted central catheters (PICC lines) and catheter tip cultures. The BacT/ALERT system was used for studying the blood culture samples. It is a quantitative blood culture system, continuously monitoring the blood for bacteria and fungus every 10 minutes. It works on colorimetric detection of carbon dioxide (CO₂) produced by the organisms inside the blood culture bottles, which is sensed by a CO₂ sensor. Positive cultures are recognized by a computer-driven algorithm that monitors both initial and increased concentrations of CO₂.

The bacterial isolates were identified and antimicrobial susceptibility testing on isolates were performed using VITEK 2 ID card and VITEK 2 AST cards respectively (bioMérieux Inc., Durham, NC, USA). It is an automated microbiology system utilizing growth-based technology.

Institutional antibiotic use policy

The institutional policy was to start all patients with febrile neutropenia on empirical antibiotic therapy with cefoperazone-sulbactam and aminoglycoside. The use of higher end antibiotics like piperacillin-tazobactum, meropenem, tigecycline, colistin, vancomycin or teicoplanin were as per the report of culture and sensitivity or when the clinical scenario deteriorated over time as per the clinician's discretion in consultation with the hospital infection control (HIC) team.

Statistical analysis

Variables studied included clinical parameters including age, co-morbidity, primary malignancy, presence of fever, number of days of intensive care unit (ICU) and hospital stays, outcomes including whether recovered or succumbed and laboratory parameters including specimen types, presence of co-infection, blood values like total leukocytic count (TLC), differential leukocytic count (DLC) and absolute neutrophil count (ANC) and antibiotic sensitivity profile. Descriptive statistics were used to elucidate the results with the use of median values, range and percentages.

Working definition of Febrile Neutropenia: Febrile neutropenia was defined as an oral temperature of \geq 38°C

on two occasions, at least one hour apart with a 12-hour period or a single temperature of $> 38.5^{\circ}$ C with an absolute neutrophil count of $\le 0.5 \times 10^{9}$ /L or $\le 1.0 \times 10^{9}$ with a predictable decline to $\le 0.5 \times 10^{9}$ /L in 24 to 48 hours.

Results

A total of 378 blood culture samples were collected from paediatric patients less than the age of 18 years during the study period. Febrile neutropenia, as per definition, was found in 252 patients (66.7%). Thus, the denominator for interpretation of results of our study is 252. There were 45 positive blood cultures (17.8%) among them. Gram-negative and gram-positive organisms accounted for (28/45) 62% and (17/45) 38% of all positive cultures respectively. The demographic and clinical characteristics of the patients are enlisted in Table 1.

Escherichia coli (39%) was the most common gram-negative isolate, followed by *Klebsiella pneumoniae* (32%), *Pseudomonas aeruginosa* (18%) and *Acinetobacter baumannii* (7%). The other organism identified was *Burkholderia cepacia* (one isolate). Coagulase-negative *Staphylococci* (CoNS) was the most common gram-positive isolate (47%) and it was followed by *Staphylococcus aureus* (35%) and *Enterococcus faecalis* (18%). Table 2 shows the details of the microorganisms isolated in the blood cultures.

Sensitivity to beta-lactam/beta-lactamase inhibitor

Table 1. Demographic and Clinical Characteristics of the Patients

Characteristics	Number/ Value (Total number = 45)			
Age				
\leq 12 months	2			
>1 year to \leq 12 years	33			
> 12 years to < 18 years	10			
Sex				
Male	25			
Female	20			
Cancer type				
Acute myelocytic leukaemia (AML)	20			
· Induction	15			
· Consolidation	3			
· Maintenance	2			
Acute lymphoblastic leukaemia (ALL)	17			
· Induction	6			
· Consolidation	6			
· Re-intensification	3			
· Maintenance	2			
Diffuse large B cell lymphoma	3			
Lymphoblastic lymphoma	1			
Burkitt lymphoma	1			
Ewing's sarcoma of bone	1			
Neuroblastoma	1			
Soft tissue sarcoma	1			

Table 2.	Organisms	Isolated	from	Blood	Cultures
----------	-----------	----------	------	-------	----------

apjcc.waocp.com

Organism	Number (Percentages)		
Gram positive organisms	17/45 (38)		
Coagulase-negative staphylococcus (CoNS)	08 (47)		
Staphylococcus aureus	06 (35)		
Enterococcus faecalis	03 (18)		
Gram-negative organisms	28/45 (62)		
Escherichia coli	11 (39)		
Klebsiella pneumoniae	09 (32)		
Pseudomonas aeuginosa	05 (18)		
Acinetobacter baumannii	02 (7)		
Burkholderia cepacia	01 (4)		

MSSA, Methicillin sensitive Staphylococcus aureus; MRSA, Methicillin resistant Staphylococcus aureus

|--|

Isolate	Aminoglycosides (%)	Carbapenems (%)	BL/BLI (%)	Colistin
Escherichia coli	6/11 (55)	4/11 (36)	2/11 (18)	9/11 (82)
Klebsiella pneumoniae	4/9 (44)	4/9 (44)	3/9 (33)	8/9 (89)
Pseudomonas aeruginosa	5/5 (100)	4/5 (80)	3/5 (60)	5/5 (100)
Acinetobacter baumannii*				
Burkholderia cepacian**				

BL/BLI, Beta lactam/ Beta lactamase inhibitor; *Only two isolates and ** only one isolate and so percentages of sensitivity not included.

							*)
Organism	ERYC	CIP	MET	GENT	VANCO	TEC	LIN
Staphylococcus aureus (n=6)	2 (33.33)	4 (66.66)	3 (50)	5 (83)	6 (100)	6 (100)	6 (100)
CoNS (n=8)	3 (37.5)	5 (62.5)	2 (25)	8 (100)	8 (100)	8 (100)	8 (100)
Enterococcus species. (n=3)	1 (33.33)	2 (66.66)	-	2 (85.71)	2 (100)	2 (100)	2 (100)

Table 4. Antibiotic Sensitivity Pattern in (%) Most of the Prevalent Gram-positive Bacteria Sensitivity (%)

*Results are expressed as a percentage of the number of isolates in each group. ERYC – Erythromycin; CIP – Ciprofloxacin; MET – Methicillin; GENT – Gentamicin; TEC – Teicoplanin; LIN – Linezolid; CoNS – Coagulase-negative Staphylococci

(BL/BLI) antibiotics like cefaperazone/sulbactam was seen in 60% of Pseudomonas isolates but the sensitivity was much less in case of *Klebsiella* species (33%) and Escherichia coli isolates (18%) (Table 3).

Sensitivity to aminoglycoside group of antibiotics was seen in all Pseudomonas isolates (100%) while 55% of *E. coli* and 44% of *Klebsiella* species were sensitive to them. Carbapenem sensitivity was noted in 80% of Pseudomonas isolates and 44% and 36% of Klebsiella species and E. coli respectively. Resistance to colistin was seen in our study. Sensitivity to colistin was noted in 89% of Klebsiella and 82% of E. coli isolates. All pseudomonas isolates were colistin sensitive. ESBL (Extended spectrum beta-lactamase) producer prevalence was 90% (18 out of 20 samples). This included 9 out of 11 isolates of E. coli and all 9 isolates of K. pneumoniae. The prevalence of CRE (Carbapenem-Resistant Enterobacterales) was 60% (12 out of 20 samples). This included 7 out of 11 isolates of E. coli and 5 out of 9 isolates of K. pneumoniae.

The incidence of methicillin resistant staphylococcus aureus (MRSA) was 50%. There was no vancomycin resistant enterococcus (VRE) in our study (Table 4).

The use of high-end antibiotics including colistin, vancomycin, tigecycline and teicoplanin was seen in five patients (11.1%) based on culture and sensitivity profile of the organisms.

The mortality rate seen in our study was 15.5% (7 patients). Out of them, five were patients with acute myeloid leukaemia, all in induction phase of treatment. The pathogens recovered in their blood cultures were Klebsiella pneumoniae (2 patients), Escherichia coli (1 patient), Enterococcus spp. (1 patient), Staphylococcus aureus (1 patient). The other two patients who died were on treatment for diffuse large B-cell lymphoma and B cell acute lymphoblastic leukaemia (ALL) on induction phase and both had Escherichia coli recovered from their blood samples.

All the patients with febrile neutropenia and positive blood culture were receiving anti-cancer drugs for their underlying malignancy and this complication resulted in treatment interruption in all of them. The use of granulocyte-monocyte colony stimulating factor (GM-CSF) is universal in patients developing neutropenia due to anti-cancer treatment, including in those who develop febrile neutropenia. Neither granulocyte transfusion nor buffy coat transfusion was done in our patients.

Discussion

We have taken the United Kingdom (UK) National Institute of Health and Care (NICE) definition of febrile neutropenia as the working definition of the condition to do our study [3]. Febrile neutropenia is one of the common complications of treatment of cancer in the paediatric age group. The incidence of this condition has been variously reported to be between 2 to 21%. It is usually seen when myelosuppressive drugs are used for the treatment of haematological or solid cancers. The contributory factors to the development of febrile neutropenia include decreased blood cell counts, marrow replacement, qualitative defects of humoral and cellular immunity, catheter-related infections and mucositis. Chemotherapy-induced mucositis compromises the gastrointestinal mucosal barrier and causes translocation of gut organisms into the bloodstream. This accounts for the gram-negative pathogens isolated in this condition [4,5,6].

The prevalence and spectrum of bloodstream infections is varied for different geographical regions [7,8,9]. The developed countries usually report a higher incidence of gram-positive organisms in blood. However, the trend seen in our region of the World is still predominantly gram-negative bacteremia. The lower use of long-term central venous catheters in developing nations compared to developed ones may be one of the reasons [10,11]. The reverse may also be true. A study was done in El Salvador, which is a developing country, comprising of 85 patients of paediatric age group who had febrile neutropenia in the study period of one year. The authors reported a higher incidence of gram-positive bacteremia (60.9%) compared to gram-negative infections (47.8%) [12]. This highlights the need of local database of the prevalent microbiological profile. In our study, we found 62% and 38% of gram-negative organisms and gram-positive organisms in the blood of the patients with febrile neutropenia with documented bloodstream infection. In a study from a tertiary care cancer centre in South India, out of 1045 blood culture samples in patients with febrile neutropenia, only 82 (7.5%) were positive. The same study showed 61% and 39% of gram-negative and gram-positive bacteremia respectively [13]. In another study from a different oncology centre in South India over a two-year period, out of 300 patients with febrile neutropenia, 15% blood culture samples had isolates and gram-negative and gram-positive organisms were identified in 58% and 40% cases respectively with 2% being fungi [14]. To the best of our knowledge, our report is the first about the microbiological profile of bloodstream infections in febrile neutropenia among cancer patients from North-East India.

A high degree of resistance was seen against betalactam/beta-lactamase inhibitor (BL/BLI) antibiotics (cefaperazone-sulbactam) in our study. This is often the first-line treatment for febrile neutropenia at our institute as well as in many other centres in India and the world. Sensitivity of aminoglycosides to *E. coli* and *Klebsiella* species was very less in our study and these are two very important pathogens. Hence, the knowledge of these resistance patterns is very important to the clinicians in deciding antimicrobial treatment correlating with the clinical course of the disease. Even colistin resistance was seen in 11% and 18% of *Klebsiella* and *E. coli* isolates respectively. The incidence of MRSA was very high (50%) in our study and this is another alarming finding. In a study from a tertiary cancer centre in Western India, out of 484 isolates that represented bloodstream infections, an 18% incidence of oxacillin resistance was noted among the *Staphylococcus aureus* isolates [15]. Fortunately, our study did not reveal any isolate of vancomycin resistant *enterococcus* (VRE).

One limitation of our study is that we did not correlate the presence or absence of malnutrition with the clinical outcomes. Studies have shown that the presence of protein-energy malnutrition (PEM) significantly increases the adverse outcomes in patients with febrile neutropenia with almost doubling of mortality rate [16]

We believe that the results of our study will add to the existing data on the epidemiology of infections in paediatric cancer patients while being treated in various cancer centres in India.

In conclusion, the knowledge of the microbiological profile and resistance patterns among patients treated for paediatric cancer with febrile neutropenia is a key factor in deciding the antimicrobial policy, preventing antibiotic resistance and counteracting adverse clinical outcomes.

References

- Collin BA, Leather HL, Wingard JR, Ramphal R. Evolution, incidence, and susceptibility of bacterial bloodstream isolates from 519 bone marrow transplant patients. Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America. 2001 Oct 01;33(7):947-953. https://doi.org/10.1086/322604
- Lyman GH, Rolston KV. How we treat febrile neutropenia in patients receiving cancer chemotherapy. Journal of Oncology Practice. 2010 05;6(3):149-152. https://doi. org/10.1200/JOP.091092
- 3. Bate J, Gibson F, Johnson E, Selwood K, Skinner R, Chisholm J. Neutropenic sepsis: prevention and management of neutropenic sepsis in cancer patients (NICE Clinical Guideline CG151). Archives of Disease in Childhood. Education and Practice Edition. 2013 04;98(2):73-75. https://doi.org/10.1136/archdischild-2013-303634
- Davis K, Wilson S. Febrile neutropenia in paediatric oncology. Paediatrics and Child Health. 2020 03;30(3):93-97. https:// doi.org/10.1016/j.paed.2019.12.002
- Barton CD, Waugh LK, Nielsen MJ, Paulus S. Febrile neutropenia in children treated for malignancy. The Journal of Infection. 2015 06;71 Suppl 1:S27-35. https://doi. org/10.1016/j.jinf.2015.04.026
- 6. Klastersky J, Naurois J, Rolston K, Rapoport B, Maschmeyer G, Aapro M, Herrstedt J. Management of febrile neutropaenia: ESMO Clinical Practice Guidelines. Annals of Oncology: Official Journal of the European Society for Medical Oncology. 2016 09;27(suppl 5):v111-v118. https://doi. org/10.1093/annonc/mdw325
- Bakhshi S, Padmanjali KS, Arya LS. Infections in childhood acute lymphoblastic leukemia: an analysis of 222 febrile neutropenic episodes. Pediatric Hematology

and Oncology. 2008 06;25(5):385-392. https://doi. org/10.1080/08880010802106564

- Prabhash K, Medhekar A, Biswas S, Kurkure P, Nair R, Kelkar R. Comparison of in vitro activities of ceftazidime, piperacillin-tazobactam, and cefoperazone-sulbactam, and the implication on empirical therapy in patients with cancer. Indian Journal of Cancer. 2009;46(4):318-322. https://doi. org/10.4103/0019-509X.55552
- Gupta A, Singh M, Singh H, Kumar L, Sharma A, Bakhshi S, Raina V, Thulkar S. Infections in acute myeloid leukemia: an analysis of 382 febrile episodes. Medical Oncology (Northwood, London, England). 2010 Dec;27(4):1037-1045. https://doi.org/10.1007/s12032-009-9330-9
- Lima MAF, Sá Rodrigues KE, Vanucci MF, Silva PLL, Baeta T, Oliveira IP, Romanelli RMC. Bloodstream infection in pediatric patients with febrile neutropenia induced by chemotherapy. Hematology, Transfusion and Cell Therapy. 2023;45(2):170-175. https://doi.org/10.1016/j. httc.2021.08.005
- Lehrnbecher T, Varwig D, Kaiser J, Reinhardt D, Klingebiel T, Creutzig U. Infectious complications in pediatric acute myeloid leukemia: analysis of the prospective multiinstitutional clinical trial AML-BFM 93. Leukemia. 2004 01;18(1):72-77. https://doi.org/10.1038/sj.leu.2403188
- Gupta S, Bonilla M, Gamero M, Fuentes SL, Caniza M, Sung L. Microbiology and mortality of pediatric febrile neutropenia in El Salvador. Journal of Pediatric Hematology/ Oncology. 2011 05;33(4):276-280. https://doi.org/10.1097/ MPH.0b013e31820ff632
- Radhakrishnan V, Vijaykumar V, Ganesan P, Rajendranath R, Trivadi G, Tenali S. Bloodstream infections in pediatric patients at Cancer Institute, Chennai. Indian Journal of Cancer. 2014;51(4):418-419. https://doi.org/10.4103/0019-509X.175360
- 14. Babu KG, Lokanatha D, Lakshmaiah KC, Suresh Babu MC, Jacob LA, Bhat GR, et al. Bloodstream infections in febrile neutropenic patients at a tertiary cancer institute in South India: A timeline of clinical and microbial trends through the years. Indian Journal of Medical and Paediatric Oncology: Official Journal of Indian Society of Medical & Paediatric Oncology. 2016;37(3):174-182. https://doi.org/10.4103/0971-5851.190352
- Prabhash K, Medhekar A, Ghadyalpatil N, Noronha V, Biswas S, Kurkure P, Nair R, Kelkar R. Blood stream infections in cancer patients: a single center experience of isolates and sensitivity pattern. Indian Journal of Cancer. 2010;47(2):184-188. https://doi.org/10.4103/0019-509X.63019
- Adejumo AC, Akanbi O, Pani L. Protein Energy Malnutrition Is Associated with Worse Outcomes in Sepsis-A Nationwide Analysis. Journal of the Academy of Nutrition and Dietetics. 2019 Dec;119(12):2069-2084. https://doi.org/10.1016/j. jand.2019.04.019

This work is licensed under a Creative Commons Attribution-Non Commercial 4.0 International License.