

Molecular Monitoring of Chronic Myeloid Leukemia in Chronic Phase (CML-CP)

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Objective: Quantification of the BCR-ABL transcript is recommended to follow-up CML patients that treated by Imatinib mesylate (IM) as a tyrosine kinase inhibitor. BCR-ABL transcripts have been recognized as a molecular marker for response to therapy in CML patients (pts). Monitoring of this marker to be more effective for identifying optimal responses and can help to inform the decision to switch to alternative therapies. Quantitative reverse transcriptase PCR (Q-PCR) of BCR-ABL1 RNA is a critical laboratory technique for accurate and sensitive monitoring of the efficiency of tyrosine kinase inhibitor therapy. The aim of our study was to analyze the molecular response (MR) in Kurdish CML patients who are treated with Imatinib.

Materials and Methods: We studied 60 blood samples from CML patients in chronic phase (CP), 36 females and 24 males, under IM treatment and monitored by Q-PCR on 12 months. The median duration of CML was 5 years (range: 1-15). The median duration of IM treatment was 4 years (range: 1-10).

Results: 40% (24 pts), 28.33% (17 pts) and 15% (9 pts) and respectively had reached early molecular response (EMR) at 1.0-2.0 log, major molecular response (MMR) at 3.0 log and deep molecular response (DMR) at 4.0-5.0 log and also undetectable BCR-ABL1 levels (CMR) were achieved in 16.67% (10 pts) at 12 months.

Conclusion: We highlighted the possibility to use of Q-PCR as a warning at diagnosis, and may use to identify patients who could benefit from a more scrupulous follow-up.

Introduction

Chronic myeloid leukemia (CML) is a hematologic stem cells disorder that determined by increased of myeloid cells in the bone marrow, spleen and peripheral blood [1, 2]. The majority of patients with CML (~ 80%) is diagnosed during the initial chronic phase (CP) and usually present with constitutional symptoms, left-shifted neutrophilic splenomegaly, and leukocytosis [3, 4]. If untreated, CML progresses to an accelerated phase (AP) and ultimately to a blast phase (BP) [3]. A sign of CML is the presence of t(9;22) (q34; q11) translocation and the resulting BCR-ABL1 gene rearrangement, which is known as Philadelphia chromosome (Ph) [1, 5]. Ph is found in more than 90% of CML patients and typically leads to p210 kDa BCR-ABL as a chimeric fusion protein [5, 6]. This fusion protein is an active tyrosine kinase that is the cause of CML disorder [1, 6]. Since the advent of BCR-ABL1 tyrosine kinase inhibitors (TKIs), patients with CML have significantly improved overall survival (OS) and lower rates of disease progressions compared with conventional therapies [7,9]. In 2001 Imatinib mesylate (IM) or Gleevec is a first TKI accepted by the US Food and Drug Administration as an anticancer that targets the adenosine triphosphate (ATP) binding

site of the kinase domain of ABL and blocks the kinase activity of BCR-ABL that is an effective treatment for chronic myeloid leukemia (CML) [10, 11]. Three levels of response in CML patients are Hematologic, cytogenetic, and molecular [12]. Complete hematological response (CHR) is defined as a standardization of the peripheral blood counts with normal peripheral blood smear, and normal spleen [13, 14]. Complete cytogenetic response (CCyR) is defined as a response to treatment of CML that absence of the Philadelphia chromosome in the marrow metaphases and Molecular response (MR) is defined as measurement of the reduction of BCR-ABL1 fusion transcripts in the quantitative RT-PCR of blood cells, by molecular methods, it is possible to follow the dynamics of minimal residual disease (MRD) over time [12, 14,16]. So quantitative reverse transcriptase polymerase chain reaction (Q-PCR) is recommended for accurate and sensitive molecular response (MR) follow-up of treated CML patients these results are expressed as a ratio of BCR-ABL on a control gene [17]. The present study was carried out to monitor the therapeutic response to IM in patients with CML in CP by BCR-ABL Q-PCR techniques.

Materials and Methods

This study was done at the laboratories in Kermanshah, Iran, between September 2014 and December 2017, 60 adult patients with CML aged 21 to 80 years old, undergoing treatment with Imatinib 400mg/day (Tabl 1). Identification of CML was determined by the existence of the BCR-ABL transcripts by RT-PCR. All CML patients identified as Chronic Phase that now on treatment were included in the study. The Chronic Phase was well-defined by the presence of < 15% blasts, < 20% basophils and < 30% blasts and also promyelocytes in both bone marrow and peripheral blood; platelet counts of at least 1×10^5 per cubic millimeter [17-19]. In order to measure BCR-ABL1 transcript levels, 3 mL of peripheral blood sample was collected in ethylenediaminetetraacetic acid (EDTA)-treated microtube. On the same day, the RNA extraction was done by using RNX-Plus kit (Sinaclon, Iran). Extracted RNA was verified by agarose (1%) gel electrophoresis and quantity and quality of extracted RNA were analyzed using a Nanodrop spectrophotometer with the measurement of the absorbance at a wavelength of 260 nm and the ratio of absorbance at wavelengths of 260 and 280 nm (260/280), respectively for cDNA synthesis. In order to synthesize of cDNA was used two-step RT-PCR kit (Vivantis, Malaysia). One μg RNA was reverse transcribed with 10 U/ μL MMLV, in 1x RT buffer, 25 ng/ μL random hexamer primer, 25 μM dNTP, 0.01 M DTT, and 2 U/ μL RNase inhibitor at 75°C for two min, 42°C for one hr, and 75°C for 10 min. with using of allele-specific primers for p190 and p210, as previously designated by van Dongen *et al* BCR-ABL1 transcripts were detected by PCR (20). PCR was done in a total volume of 25 μL of a reaction mixture comprising 2.5 μL of 10X PCR buffer, 1 μL of 50 mM MgCl_2 , 1 μL of 10 mM dNTPs, 2 μL of 10 μM of forward and reverse primers, 18 μL of H_2O and 0.5 μL of 5U/ μL *Taq* DNA polymerase. The program that was considered for thermal cycler including: initial denaturation for 5 min at 94°C, followed by 35 cycles of denaturation for 45 seconds at 94°C, annealing as 50 seconds at 58°C, extension 40 seconds at 72°C and final extension on 7 minutes at 72°C. The PCR products were electrophoresed on a 2.5% Agarose gel with gel red. Pretreatment BCR-ABL1 transcript levels were measured by RT-PCR and after the beginning of IM therapy BCR-ABL1 transcript levels were measured at 12 months, the results were calculated as a percentage of the ratio of *BCR-ABL/ABL* copy numbers. Molecular response to therapy with IM was defined as: Major molecular response (MMR): if their *BCR-ABL/ABL* ratios showed a reduction to 0.1% ($\leq 3\log$) or 0.005% ($\leq 4.5\log$); deep molecular response (DMR): if their *BCR-ABL/ABL* ratios showed > 4 or 4.5 log reduction. Patients achieving a DMR are possible candidates for treatment free remission trials; complete molecular response (CMR): according to the International Scale (IS), BCR-ABL transcripts not detectable, from a standardized baseline [21]. Also according to the definitions of failure of and suboptimal response to Imatinib suggested by the European Leukemia Network (ELN),

these responses were calculated at 12 months (Tabl 1) [22].

	Hematologic Response		Molecular Response
		Cytogenetic	



		Response	
	3months	Not full	None (ph+>95%)
Failure			
	3months		
Warning	6months		
	12months		

Table 1. Definition of the response to TKIs as first-line treatment recommended by ELN.

Statistical analyses

At diagnosis CML, the Sokal index and Hasford risk score were calculated [23, 24]. All of the statistical analyses were performed using Excel and R software. A database was created in MS Excel Statistical significance was considered at the p < 0.05 level.

Results

The available demographic and clinical parameters of the CML patients are presented in Tabl .

Variable	CML patients (N=60)
Median age, (range), years	41 (21-80)
Sex (female/male)	60 (36/24)
Median CML duration, years	5 (1-15)
The median duration of IM treatment, (range)	4 (1-10)
	82 (10-630)
	10.9 (6-15.6)

Platelets (10/L), median (range)	393 (13-1280)
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Table 2. Characteristics of CML patient

Overall, 60 CML patients with chronic phase were included in our study. There were 24 (40%) male and 36 (60%) female patients. The median duration of CML was 5 years (range: 1 year-15 year) and the median duration of IM treatment was 4 years (range: 1 year-10 year). Sokal Score for 52 patients (86.6%), 4 (6.7%) and 4 (6.7%) were low, intermediate and high, respectively, but Hasford (Euro) Score for all patients was low. All of the 60 patients were positive for BCR-ABL at diagnosis time and BCR-ABL: ABL levels in these patients were above and various at baseline. As shown in Fig.1, in our study after 12 months from Imatinib therapy, BCR-ABL: ABL ratio was monitored at baseline.

Figure 1: Competitive quantitative PCR values at 12 months on IM therapy by the molecular response

The rates of MMR and DMR at 12 months were 28.33% and 13.33%, respectively. Also, CMR levels were achieved in 16.66% at 12 months (Tabl 3).

Log reduction BCR-ABL/ABL ratio	Frequency	Percent
1 <log reduction ≤2 (EMR)	24	40%
3 log reduction (MMR)	17	28.33%
4 ≤ log reduction ≤5 (DMR)	9	15%
CMR= Zero	10	16.67%
Total	60	100

Table 3. Log reduction of BCR-ABL/ABL ratio at 12 months while on 400 mg/day imatinib ($\chi^2= 3.019$; $P<0.05$).

Of the 60 patients included in the study, 58.33% finally achieved optimal response. Of the others, 20 (33.33%) and 5 (8.33%) patients respectively had treatment failure and suboptimal response according to the definitions ELN.

Discussion

For the purpose of targeted therapy for patients with chronic myeloid leukemia (CML), monitoring is vital for optimizing treatment and affordable results. Molecular monitoring in the CML is a noninvasive method to predict outcome and treatment guide decisions. Since 2001 the Imatinib treatment has been used as a front-line therapy in CML patients [7]. This type of treatment is associated with high rates of MMR although a higher rate of CMR (undetectable BCR-ABL) has been reported in patients getting prolonged Imatinib therapy, significant numbers of patients do not achieve MMR, have to be at risk for disease progression [25, 26]. Increasing levels of BCR-ABL1 RNA indicate the progression of the disease, so the molecular response became an important analytical factor to predict the outcome of the patients treated with TKIs [2]. It is now widely accepted Q-PCR has been used to monitor MRD in CML patients [27].

In our study, the response to treatment is evaluated in patients that received 12 months Imatinib by monitored the BCR-ABL/ABL ratio, and the levels of MMR, DMR, and CMR were 28.33%, 15%, and 35%, respectively. These results are relatively comparable with the results reported by Ozatli et al. (2010) that in 77 Turkish CML patients MMR and CMR rates were 58.5% and 32.5% at 12 months Imatinib treatment, respectively [28].

But these results are fewer than a Study that performed by Bixby (2009) in 204 subjects were reported 50.1% MMR at one-year treatment [29].

We supposed that according to different groups of the patients, response rates can be changeable because of gene polymorphisms in different countries and these results might be due to a small sample size. In the future, we hope that will have more samples, so results will be more accurate in representing the population.

The number of patients who achieve a DMR may enable to stop treatment and proper monthly qPCR monitoring [30]. In the case of a contradiction between MR results at prior time points the pathologist should be recommended resampling for validation of results before any decision is taken [25].

suboptimal response and failure treatment, according to the latest ELN recommendations should be considered for a change of treatment [31]. In the present study, 33.33% of patients didn't the response to 400 mg/day IM so in this case we are faced with two solutions first, change in dose of Imatinib second, a mutation analysis in the kinase domain of BCR-ABL. More than 60 various mutations are identified to be associated with resistance to Imatinib. If Y253H, E255K/V, V299L, T315I, F317L, F359V mutations are identified, the pathologist should indicate which TKI is probably to be more effective against that specific mutation because of resistance is thus associated with a greater risk of disease progression [25].

Though the advent of TKIs has improved patient outcomes, CML is not easy to manage, but the quantitative determination of residual disease with Q-PCR is a reliable method for monitoring molecular response that provides critical analytical data for the management of CML patients which is treated with TKI therapies.

So we conclude that regular molecular monitoring (every 6 or 12 months) of BCR-ABL1 by RT-qPCR provides essential information for evaluating responses as well as for forecasting progression-free survival or relapse during TKI therapy in patients with CML. In this study, A \geq 3-log reduction in 75% patients at 12 months was attained. Patients who achieve \geq 1-log reduction at 6 months and \geq 2-log reduction at 12 months are more probably to successively reach MMR [32]. So the early trend in the BCR-ABL/ABL ratio might be clinically useful for the early identification of patients who are destined to achieve an MR on Imatinib.

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