

The Association between PM_{2.5} Exposure and Hippocampal Volume: A Systematic Review

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Background: Existing air quality is decreasing, as evidenced by the increase in air pollution. Air pollution does not only affect the respiratory system, but also affecting the nervous system, and furthermore causing impaired cognitive function that can be predicted through the image of the hippocampus.

Objective: This study wanted to determine the significance of the relationship between PM_{2.5} (Particulate Matter) pollutant exposure and hippocampal volume in adults.

Method: This research is a PRISMA 2020 based systematic study using Google Scholar, PubMed, and Proquest as databases. Research inclusion criteria were studies with subjects over 19 years old, using MRI techniques, published in English, having sufficient data for extraction.

Result: There are 5 studies from 2015 to 2020 which stated that there was no statistically significant relationship between PM_{2.5} pollutant exposure and hippocampal volume (n = 5) (P-value > 0.05, 0.71, 0.8, 0.32), and the study obtained significant results (n = 1) (P-value < 0.005). Discussion: Although the results of the study did not prove a significant difference in hippocampal volume, several recent theories regarding hippocampal neurogenesis in adults are able to support these results.

Conclusion: From this study, it was not proven that there was a significant relationship between PM_{2.5} pollutant exposure and hippocampal volume.

Introduction

According to IQAir website, there was an increase in the concentration of particulate matter (PM_{2.5}) by more than 50% in 2018 [1]. Chronic exposure to air pollution causes multisystem disturbances [2]. Regardless of the known etiology, several studies suggest that exposure to air pollution can have a negative impact on the environment, adverse effects on the human brain, including cognitive function [2,3]. Several previous studies have strongly associated PM_{2.5} with decreased total cerebellar volume (TCBV), but few results have been obtained regarding PM_{2.5} with hippocampal volume. From the literature search, we get different results statistically, therefore it is necessary to carry out a systematic review to this topic.

Particulate Matter (PM) which categorized as ambient (outdoor) pollutants is divided into 3 types based on its size: coarse (PM₁₀), fine (PM_{2.5}), and ultrafine (PM_{0.1}). Produced from coal burning, power plant waste, motor vehicle fumes, and forest fires [4]. Based on the literature search of this study, neuronal damage due to PM exposure can occur through two pathways: mediated by hypoxia and mediated by blood brain barrier damage. Theoretically, hypoxia will cause hippocampal atrophy.

From the results of research on hypoxia in neonates, it is proven that hypoxia, hippocampal atrophy, and memory impairment are one-way events that occur sequentially through activation of HIF-1 α (hypoxia inducible factor 1 alpha), degradation of MAP-2 protein, formation of oxidative stress, and neuronal apoptosis [4-11]. In another study, it was explained that PM exposure caused several changes in the hippocampus of mice, such as impaired methionine-glutathione metabolism which indicated an imbalance of glutathione redox, which was characterized by the formation of oxidative stress, then hippocampal neuroinflammation, and an increase in A β levels [12].

Methods

This study is previously registered in PROSPERO with registration ID CRD42021264296. Primary studies used obtained through a literature search using Google Scholar, PubMed, and ProQuest databases. The keywords searched through Mesh-Term are "air pollution", "hippocampal volume", "magnetic resonance imaging", and "adult" entered into the 3 databases used. We used advanced search and limitations for English and Indonesian studies, free full text, and publications in the previous 5-10 years. We selected studies based on PRISMA 2020 guidelines and pre-defined inclusion criteria, in the form of studies with subjects over 19 years of age, using MRI techniques, published in English, and sufficient data for extraction. Study screening was carried out based on the title, abstract, and full text review. To minimize bias, we used two valid instruments, CASP and AXIS based on the research method used in each study. The studies that qualified the inclusion criteria are then carried out with data extraction: population characteristics, pollutant measurement and analysis methods, as well as imaging methods and hippocampus image analysis.

Results

From the results of a literature search using the PRISMA 2020 method (Figure 1), studies using cross-sectional method (n = 3) and longitudinal cohort (n = 3) were obtained.

Figure 1. Literature Search Result Using PRISMA Flowchart 2020.

Characteristics of the studies we found are study subjects aged ≥ 60 years (n = 6), excluding subjects with history of neurological disease (n = 4). Measurement of pollutant concentration by BME (n = 2), spectroradiometer (n = 1), spatiotemporal statistical model (n = 1), and national prediction model (n = 1). Imaging of the hippocampus with T1-weighted MRI (n = 1), T2-weighted

MRI (n = 2), and T1-weighted MPRAGE MRI (n = 3). The data obtained were then processed statistically to determine the association of the two variables. Spearman test (n = 2) and cross-validation test (n = 3) are used to determine the validity of the pollutant concentration data, while the statistical tests used to assess the correlation between PM2.5 exposure and a decrease in hippocampal volume include linear regression, logistic regression test, and chi-square test (Table 1).

Study	Age of Subject	Inclusion Criteria	Pollutant Concentration Measurement		Hippocampal Imaging		P-value
			Method	Statistical Analysis	Method	Statistical Analysis	
Chen et al [2015][13]	65-80	No history of dementia	BME	r = 0,90	T1-weighted volumetric MRI scans for 7-10 years follow up	Automatic computer-based template warping	0,8
Wilker et al [2015][14]	≥60	No history of dementia, stroke. Attended 7 examinations.	Spectroradiometer-meter	r = -0,15	MRI T2-weighted double spin-echo	Manual delineation	<0,005
Casanova et al [2016][15]	65-80	Not mentioned	BME	R ² = 0,74	MRI T2-weighted spin echo	Regional Analysis of Volumes Examined in Normalized Space (RAVENS)	>0,05
Power et al [2018][16]	Mean = 76±5	No history of head surgery or radiation therapy, multiple sclerosis, brain tumor, and attended 5 examinations.	Spatiotemporal statistical model	R ² = 0,50 - 0,83	MRI T1-weighted 3D volumetric magnetization-prepared gradient echo (MPRAGE)	Semi-automatic software Freesurfer	0,71
Cho et al [2020][17]	Mean = 67,3	No history of neurological disease such as dementia and has had imaging examination.	National prediction model	R ² = 0,45	MRI T1-weighted 3D MPRAGE	Semi-automatic software Freesurfer	0,32
Hedges et al [2020][18]	Mean = 62,15	Not mentioned	Data collected from UK Biobank		MRI T1-weighted 3D MPRAGE	Automatic device	>0,05

Table 1. Characteristics of Studies Included in Systematic Review.

Note, BME (Bayesian Maximum Entropy)

Not all studies used the same variable measurement method, and there are studies that used manual delineation method to analyze the results of MRI images (n = 1). Since the studies are not homogeneous enough, meta-analysis cannot be carried out. The results of the studies that have been qualified are statistically significant (n = 1) and not significant (n = 5).

Discussion

Based on the systematic study conducted, it was stated that there was no statistically significant relationship between PM_{2.5} exposure and hippocampal volume as measured by MRI results (n = 5). Mean while, another study stated that there was a significant relationship with p-value <0.05 (n = 1). Based on the theory that has been reviewed, exposure to pollutants can induce acute hypoxia, but there is still an unresolved controversy regarding the effects of acute hypoxia on hippocampal neuron cells. This theory is opposed by a study [19] which states that acute hypoxia can support hippocampal neurogenesis in adults [19] (Figure 2).

Figure 2. Adult Hippocampal Neurogenesis.

The process of hippocampal neurogenesis is regulated by the transcription factor Sox 2, expressing Sonic hedgehog (Shh) which then promotes type 1 cell proliferation. Hippocampal neurogenesis has 4 phases, namely: precursor cells, early survival phase, post-mitotic phase, and late survival phase [20]. Precursor cells are located in the sub granular zone of the hippocampus and are divided into two, neural stem cells (NSC) which consists of type 1 and type 2A cells, and intermediate neuronal progenitor (INP) which consists of type 2B and type 3 cells [21]. Precursor cells lasted for three days, during which asymmetric division of type 1 cells, which were glial cells with triangular bodies, functioned for the expression of nestin and glial fibrillary acidic protein (GFAP). Type 1 cell division will produce type 2 cells, which are also called transient cells that have the ability to migrate and proliferate, which can be further identified by the presence or absence of the expression of doublecortin (DCX). Type 2A cells do not express DCX (DCX negative), whereas type 2B cells can express DCX, which then differentiate into type 3 cells. Type 3 cells do not express nestin, however it expresses DCX and polysialylated neuronal cell adhesion molecule (PSA-NCAM). The INPs consisting of cells type 2B and 3 together then express the transcription factor T-box brain protein (Tbr2). The newly formed cells then enter the post-mitotic phase, this is characterized by the expression of the neuron nucleus (NeuN) and calretinin. Furthermore, the number of neuroblasts that have been formed will decrease due to the apoptotic process that occurs, and only 20% of neuroblasts will join the existing network of neurons [20]. There are two critical periods: the first and second critical periods. The first critical period is the transition period for Tbr2 until there is an increase in DCX expression. Neuroblasts that have been expressed will then enter the second critical period, transitioning into immature neuron granules, merging into the existing granule layer network, finally developing into mature neuron granules [19]. In addition, an experimental study in animals also found that acute hypoxia that occurs during the first critical period can increase the number of new neurons (birth-dated neurons) through HIF-1 activation which will then help neuron survival by initiating neuroprogenitor production. Further detailed mechanism of the pathway for neuroprogenitor initiation by HIF-1 is still unknown. Further research is needed [19]. By acknowledging this theory, the results of the five studies that were not significant could be explained. In conclusion, although PM_{2.5} is considered harmful, exposure to PM_{2.5} is not significantly associated with changes in hippocampal volume in adults. Further research should be focused on systematic review update and meta-analysis of smaller types of pollutants such as PM_{0.1}.

Acknowledgements

Conflict of Interest

The author has no conflict of interest to declare.

Funding

The authors declare that no funds, grants, or other support were received during the preparation of

this manuscript.

Author Contributions

The study conception and design were carried out by Yopi Simargi. Data collection, analysis, and interpretation, and first draft of the manuscript were performed and written by Elizabeth Feloni Lukito. Kevin Tandarto, Maureen Miracle Stella, Ignatius Ivan, Harvey Sudharta, Gilbert Golahi, Yuda Turana, Bryany Titi Santi, and Yanto Budiman commented on previous versions of the manuscript. All authors read and approved the final manuscript.

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